**TEST MENU**

Expected Values are final test results deemed consistent with CDC vaccination recommendations, nationally recognized reference ranges and standard values found in certain populations for clinical and non-clinical testing.

<table>
<thead>
<tr>
<th>TEST</th>
<th>DESCRIPTION</th>
<th>TRANSPORT KIT?</th>
</tr>
</thead>
</table>
| **ANTIBIOTIC SUSCEPTIBILITY (DISC DIFFUSION)** | **Description:** Kirby Bauer disc diffusion antimicrobial testing approved by CLSI or CDC (up to 12 antimicrobials)  
**Specimen:** Pure isolate in supportive environment/media  
**Instructions:** Call to confirm availability of antimicrobials and suitability of organism for testing. Transport at ambient conditions. Tubed media must be used to transport by common carrier.  
**Expected value:** Susceptible, Resistant, or Intermediate of antimicrobial | No             |
| **ANTIBIOTIC SUSCEPTIBILITY (MINIMUM INHIBITORY CONCENTRATION)** | **Description:** Minimum Inhibitory Concentration (MIC) antimicrobials using E-test technology (per antimicrobial)  
**Specimen:** Pure isolate in supportive environment/media  
**Instructions:** Call to confirm availability of antimicrobials and suitability of organism for testing. Transport at ambient conditions. Tubed media must be used to transport by common carrier.  
**Expected value:** Value of antimicrobial in ug/ml | No             |
| **ARBOVIRUS (WNV, SLE, EEE)**                 | **Description:** Detection of West Nile virus (WNV), St. Louis encephalitis virus (SLE), and eastern equine encephalitis virus (EEE) IgM antibodies by antibody capture ELISA and IgG antibodies by ELISA  
**Specimen:** 1-2 ml serum; at least 1 ml CSF  
**CSF:** Collect within the first 14 days following disease onset and refrigerate at 2-8°C until transport.  
**Serum:** Paired acute-phase (collect as early as possible after onset of illness) and convalescent phase (collect >/= 8 days after clinical onset) and refrigerate at 2-8°C. Acute serum and CSF should be sent as soon as possible to the PHL for testing.  
**Expected Value:** Negative for WNV, SLE, and EEE antibodies | Yes            |
| **ARSENIC, TOTAL**                            | **Description:** Quantitation of urinary arsenic by inductively coupled plasma mass spectrometry (ICP MS) Dynamic Reaction Cell  
*Specimen:* Urine: 20 ml in 50 ml polypropylene centrifuge tube  
*Instructions:* Refrain from seafood consumption for 3 days prior to urine collection. Freeze sample at < or = -20°C; if not possible, store sample at < or = 4°C and transport to PHL at < or = 4°C as soon as possible  
**Expected Value:** <50 µg/L | No             |
| **BACTERIAL CULTURE**                         | **Description:** Complete identification of aerobic or anaerobic bacterial organisms including grouping or typing where appropriate  
**Specimen:** Aseptically collected clinical specimen  
**Instructions:** Transport at temperatures and atmosphere to maintain viability of organisms. Call PHL for further instructions if necessary  
**Expected Value:** Absence or presence of bacterial organism(s) |
<table>
<thead>
<tr>
<th>TEST</th>
<th>DESCRIPTION</th>
<th>TRANSPORT KIT?</th>
</tr>
</thead>
</table>
| BACTERIAL IDENTIFICATION         | **Description:** Complete identification of bacterial isolate includes grouping or typing where appropriate.  
**Specimen:** Pure isolate in supportive environment/media.  
**Instructions:** Transport at room temperature in necessary atmosphere for maintenance of viability. Tubed media must be used to transport by common carrier.  
**Expected Value:** Genus and species of organism                                                                                     | No            |
| BLOOD PARASITE, MICROFILARIAE, MALARIA, BABESIA | **Description:** Giemsa stained smears, thick and thin, Knott’s concentration for microfilariae  
**Specimen:** Whole blood: Preserved with EDTA anticoagulant  
Smears: Peripheral blood from fingerstick or EDTA preserved blood; stained or unstained  
**Note:** Thick smears cannot be performed without preserved EDTA specimen  
**Instructions:** Transport at ambient conditions. Deliver as soon as possible as EDTA specimens >3 days from collection are unacceptable for testing. It is recommended that thin smears be prepared within one hour of collection. Once dried, thin smears should be fixed in methanol before transport.  
**Expected Value:** No parasites seen                                                                                           | No            |
| CHIKUNGUNYA                      | **Description:** Detection of Chikungunya virus using Real-Time RT-PCR Assay  
**Specimen:** 1 ml serum  
**Instructions:** Transport specimens to the laboratory at 2–8°C as soon as possible. If a delay greater than 24 hours is expected before specimens can be submitted to the laboratory, the serum should be separated and stored at refrigerated temperature. For long-term storage, serum samples for molecular diagnosis should be stored frozen (at −20°C for short-term storage or at −70°C)  
**Expected Value:** Chikungunya virus RNA not detected                                                                                       | Yes           |
| CHLAMYDIA TRACHOMATIS -AMPLIFIED TEST | **Description:** A Strand Displacement Amplification (SDA) screening test for the detection of Chlamydia trachomatis  
**Note:** Not to be used in medico-legal situations (see Chlamydia culture and Gonorrhea culture)  
**Specimen:** Male: Urethral swab; rectal swab; urine  
Female: Endocervical swab; rectal swab; urine  
**Instructions:** Swab specimens - may be stored and transported to the PHL at 2-27°C. Swab specimens must be tested within 6 days of collection  
Urine - 15-60 ml first-catch collected in a urine processing tube (UPT tube) containing preservative (patient should not have urinated for at least 1 hour prior to specimen collection). UPT specimens must be tested within 30 days of collection.  
**Expected Value:** Negative                                                                                                               | Yes – Request SDA swabs or UPT tubes |
| CHLAMYDIA TRACHOMATIS - CULTURE  | **Description:** Isolation of Chlamydia trachomatis from clinical specimen  
**Note:** Culture is the only C. trachomatis test useful in medico-legal situations  
**Specimen:** Cotton-tipped swabs (DO NOT use swabs with calcium alginate or wood shafts); aspirate or biopsy in universal transport media. Acceptable specimens include: cervical swab; conjunctival swab; epididymis aspirate; fallopian tube swab or biopsy; lymph node aspirate; middle ear aspirate; respiratory tract swab, aspirate or biopsy; urethral swab  
**Instructions:** Refrigerate at 2-8°C until transport. Use cold pack when transporting. Deliver to PHL within 24 hours; contact PHL prior to transport  
**Expected Value:** No evidence of Chlamydia trachomatis in culture                                                                         | Yes           |
<table>
<thead>
<tr>
<th>TEST</th>
<th>DESCRIPTION</th>
<th>TRANSPORT KIT?</th>
</tr>
</thead>
</table>
| CRYPTOCOCCAL ANTIGEN                     | **Description:** Latex agglutination test for Cryptococcal polysaccharide  
**Specimen:** 1 ml CSF or 2 ml serum  
**Instructions:** Refrigerate until transported, freeze if delay in transporting >72 hours. Transport at ambient conditions.  
**Expected Value:** Negative for Cryptococcal Antigen |
| CRYPTOSPORIDIUM/ GIARDIA DFA             | **Description:** Monoclonal direct fluorescent antibody (DFA) test on concentrated fecal specimen  
**Stool specimen, duodenal fluid or mucosal biopsy preserved in 10% formalin, sodium acetate-acetic acid formalin, Streck tissue fixative (STF) or Ecofix.  
**Expected Value:** None seen |
| CYCLOSPORA/ ISOSPORA/ SARCOCYSTIS         | **Description:** Direct and concentrated wet mounts, modified acid-fast stain, fluorescent MAF stains, autofluorescence  
**Stool specimen preserved in 10% formalin, sodium acetate-acetic acid formalin (SAF), Streck tissue fixative (STF) or Ecofix.  
**Expected Value:** No Cyclospora, Isospora, or Sarcocystis detected |
| EASTERN EQUINE ENCEPHALITIS VIRUS (EEE)  | See Arbovirus Serology Panel                                                                                                                                                                                                 | Yes           |
| ENTERIC SCREEN (LIMITED)                 | **Description:** Testing for *Salmonella* and *Shigella* sp. only  
**Specimen:** Stool specimen in fecal transport (Cary Blair or equivalent).  
**Instructions:** Transport at ambient conditions; must receive within 3 days of collection. Refrigerate if delay in transport >24 hours.  
**Expected Value:** No *Salmonella* isolated  
No *Shigella* isolated |
| ENTERIC SCREEN (FULL SCREENING)          | **Description:** Testing for *Salmonella, Shigella, Campylobacter, Aeromonas, Plesiomonas, Enterohemorrhagic E. coli, Yersinia sp. (complete identification if found), Shiga-like toxins*  
**Note:** Screen for enterotoxigenic *Staphylococcus aureus, Bacillus cereus*, and *Clostridium perfringens NOT* included in this screen. See Enteric Special Culture.  
**Specimen:** Stool specimen in fecal transport (Cary Blair or equivalent).  
**Instructions:** Refrigerate if delay in transport >24 hours.  
No *Salmonella, Shigella, Campylobacter, Aeromonas, Plesiomonas,*  
**Expected Value:** Enterohemorrhagic *E. coli, or Yersinia* isolated |
| ENTERIC SCREEN (SPECIAL)                 | **Description:** Testing for *Staphylococcus aureus, Bacillus cereus, Clostridium difficile culture, Clostridium perfringens*  
**Specimen:** Stool specimen  
**Instructions:** Call PHL at 603-271-4661, for specimen and transport instructions  
No *Staphylococcus aureus, Bacillus cereus, Clostridium difficile, or Clostridium perfringens* isolated |

**DISCONTINUED 2016**
<table>
<thead>
<tr>
<th>TEST</th>
<th>DESCRIPTION</th>
<th>TRANSPORT KIT?</th>
</tr>
</thead>
<tbody>
<tr>
<td>ENTEROVIRUS CULTURE</td>
<td>See Viral Culture</td>
<td>Yes</td>
</tr>
<tr>
<td>EPIDEMIOLOGY STUDY</td>
<td>(Isolate or specimen)</td>
<td>No</td>
</tr>
<tr>
<td></td>
<td>Description: Complete identification of bacterial/parasitic isolate; includes grouping or typing where appropriate</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Specimen: Pure isolate in supportive environment/media or original specimen</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Instructions: Transport at temperature in atmosphere for maintenance of viability</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Genus and species of organism</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Expected Value: Recovery of any of these organisms must be reported to the NH Bureau of Disease Control and Prevention at 603-271-4496.</td>
<td></td>
</tr>
<tr>
<td></td>
<td>NOTE: Call the NH PHL for further information 603-271-4661</td>
<td></td>
</tr>
<tr>
<td>FOOD CONTAMINANTS -GENERAL SCREEN (ADULTERATION OF TAMPERING)</td>
<td>Description: General method to examine foods and beverages for small and medium molecular weight toxic contaminants</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Specimen: At least 10 grams (if solid) or 10 ml (if liquid) of sample</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Instructions: Contact Bureau of Disease Control and Surveillance (603-271-4596), or law enforcement. Refrigerate sample in closed container.</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Expected Value: None</td>
<td></td>
</tr>
<tr>
<td>FUNGAL CULTURE WITH IDENTIFICATION</td>
<td>Description: Mycologic culture for yeasts, filamentous fungi and aerobic actinomycetes</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Specimen: Fluid, tissue, or secretions: Send in sterile container. Keep moist if necessary with sterile water, sterile saline, or brain heart infusion (BHI) broth. Blood: 10 ml whole blood in SPS tube. Swabs are unacceptable.</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Instructions: Transport at ambient conditions. Hold at 2-8°C. Specimens unacceptable &gt;7 days from collection.</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Expected Value: No fungus isolated</td>
<td></td>
</tr>
<tr>
<td>FUNGAL ISOLATE IDENTIFICATION</td>
<td>Description: Identification of fungi to genus; species level if appropriate isolate (preferably pure) on supportive media</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Specimen: Samples must be in tubed media for common carrier transport. Transport and hold at ambient conditions.</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Expected Value: Genus identification of organism</td>
<td></td>
</tr>
</tbody>
</table>

NH DHHS, Division of Public Health Services 12/9/15 Page 4 of 14
<table>
<thead>
<tr>
<th>TEST</th>
<th>DESCRIPTION</th>
<th>TRANSPORT KIT?</th>
</tr>
</thead>
<tbody>
<tr>
<td>GIARDIA</td>
<td>See Cryptosporidium/Giardia</td>
<td>Yes</td>
</tr>
</tbody>
</table>
| GONORRHEA - AMPLIFIED TEST               | **Description:** A Strand Displacement Amplification (SDA) screening test for the detection of *Neisseria gonorrhoeae*  
|                                          | **Note:** Not to be used in medico-legal situations (see Gonorrhea culture) |                |
|                                          | **Specimen:**  
|                                          | Male: Urethral swab; urine                                                 | Yes Request SDA swabs or UPT tubes |
|                                          | Female: Endocervical swab; urine                                           |                |
|                                          | **Instructions:** Swab specimens - may be stored and transported to the PHL at 2-27°C. Swab specimens must be tested within 6 days of collection.  
|                                          | Urine - 15-60 ml first-catch collected in a urine processing tube (UPT tube) containing preservative (patient should not have urinated for at least 1 hour prior to specimen collection). UPT specimens must be tested within 30 days of collection. |                |
|                                          | **Expected Value:** Negative                                                |                |
| GONORRHEA - CULTURE                      | **Description:** Isolation and identification of *Neisseria gonorrhoeae*. All isolates confirmed as *N.gonorrhoeae* are tested for resistance to Penicillin, Tetracycline, Spectinomycin, Ciprofloxacin, Ceftriaxone and Azithromycin  
|                                          | **Specimen:** Note: Culture is the only gonorrhea test useful in medico-legal situations  
|                                          | Inoculated Jembec, Martin-Lewis, Modified Thayer Martin, NYC, Thayer Martin plate, or CA enriched (GC base)  
|                                          | **Instructions:** If possible, incubate 18-24 hours at 37°C before transporting to laboratory. Otherwise, plate can remain at room temperature for 16-18 hours until transported. |                |
|                                          | **Expected Value:** No *Neisseria gonorrhoeae* isolated                    | Yes           |
| GONORRHEA - ISOLATES FOR CONFIRMATION    | **Description:** Identification and confirmation of suspect *N.gonorrhoeae* isolates. All isolates confirmed as *N.gonorrhoeae* are tested for resistance to Penicillin, Tetracycline, Spectinomycin, Ciprofloxacin, Ceftriaxone and Azithromycin  
|                                          | **Specimen:** Isolate on supportive media                                  | No            |
|                                          | **Instructions:** Transport at room temperature in necessary atmosphere for maintenance and viability. Tubed media is necessary for common carrier transport. To be received at PHL as soon as possible. |                |
|                                          | **Expected Value:** *Neisseria gonorrhoeae* isolated with antibiotic susceptibilities |                |
| HEMORRHAGIC *E. COLI* SCREEN/ CONFIRMATION| **Description:** Screen: Shiga-like toxin EIA performed; culture with confirmation to include biochemical identification, serogroup and serotype  
|                                          | **Specimen:** Screen: Fecal specimen in fecal transport (Cary Blair or equivalent). Confirmation: Pure isolate on supportive media  
|                                          | **Instructions:** Transport at ambient conditions; must receive within 3 days of stool collection. Refrigerate if delay in transport >24 hours. Tubed media must be used for transport by common carrier. | Yes           |
|                                          | **Expected Value:** No Hemorrhagic *E. coli* isolated                     |                |
| HEPATITIS A VIRUS ANTIBODY (Anti-HAV) IgM | **Description:** Detection of Hepatitis A IgM antibody by ELISA  
|                                          | **Specimen:** 1-2 ml serum or plasma                                        | Yes           |
|                                          | **Instructions:** Specimens may be refrigerated at 2-8°C for up to 5 days prior to transport. For longer storage, serum or plasma (separated from whole blood) may be frozen at -20°C or below prior to transport  
<p>|                                          | <strong>Expected Value:</strong> Negative for Hepatitis A Virus IgM antibody             |                |</p>
<table>
<thead>
<tr>
<th>TEST</th>
<th>DESCRIPTION</th>
<th>TRANSPORT KIT?</th>
</tr>
</thead>
</table>
| HEPATITIS A VIRUS ANTIBODY (Anti-HAV) TOTAL | **Description:** Detection of total antibody to Hepatitis A Virus by ELISA  
**Specimen:** 1-2 ml serum or plasma  
**Instructions:** Specimens may be refrigerated at 2-8°C for up to 5 days prior to transport. For longer storage, serum or plasma (separated from whole blood) may be frozen at -20°C or below prior to transport  
**Expected Value:** Negative for Hepatitis A Virus total antibody | Yes            |
| HEPATITIS B CORE ANTIBODY (Anti-HBc) IgM | **Description:** Detection of Hepatitis B total core antibody by ELISA  
**Specimen:** 1-2 ml serum or plasma  
**Instructions:** Specimen should be collected within 1 month of onset of symptoms. Refrigerate at 2-8°C for up to 1 day prior to transport. If longer storage is required, serum or plasma (separated from whole blood) may be frozen at -20°C or below  
**Expected Value:** Negative for Hepatitis B total core IgM antibody | Yes            |
| HEPATITIS B SURFACE ANTIBODY (Anti-HBs)  | **Description:** Detection of Hepatitis B surface antibody by ELISA  
**Specimen:** 1-2 ml serum or plasma  
**Instructions:** Specimen may be refrigerated at 2-8°C for up to 1 day prior to transport  
**Expected Value:** Positive or Negative for Hepatitis B surface antibody | Yes            |
| HEPATITIS B SURFACE ANTIGEN (HBsAg)      | **Description:** Detection of Hepatitis B surface antigen by ELISA  
**Specimen:** 1-2 ml serum or plasma  
**Instructions:** Specimen should be collected 2 to 12 weeks after onset of symptoms or whenever a chronic infection is suspected. Refrigerate at 2-8°C for up to 7 days prior to transport. For longer storage, serum may be stored at -20°C or below. Positive results will be reflexed to Hepatitis B Core Antibody IgM.  
**Expected Value:** Negative for Hepatitis B surface Antigen | Yes            |
| HEPATITIS C (HCV) ANTIBODY SCREEN        | **Description:** Detection of HCV antibody by ELISA  
**Specimen:** 1-2 ml serum or EDTA, heparinized, or citrated plasma (not heat-treated)  
**Instructions:** Specimens may be refrigerated at 2-8°C for up to 6 days. For longer storage, serum or plasma (separated from whole blood) may be frozen at –20°C or below (not freeze-thaw) prior to transport  
**Expected Value:** Negative for Hepatitis C Antibody | Yes            |
| HEPATITIS C (HCV) AMPLIFIED METHOD       | **Description:** Detection of HCV RNA by RT-PCR  
**Specimen:** 1-2 ml serum or EDTA, heparinized, or citrated plasma (not heat-treated)  
**Instructions:** Specimens may be refrigerated at 2-8°C for up to 2 days. For longer storage, serum or plasma (separated from whole blood) may be frozen at -20°C or below (not freeze-thaw) prior to transport  
**Expected Value:** HCV RNA not detected | Yes            |
<table>
<thead>
<tr>
<th>TEST</th>
<th>DESCRIPTION</th>
<th>TRANSPORT KIT?</th>
</tr>
</thead>
</table>
| HEPATITIS C VIRUS (HCV) GENOTYPING       | **Description:** Genotyping by sequence analysis of the 5'UTR region of the HCV genome  
**Specimen:** 1-2 ml serum or EDTA, heparanized, or citrated plasma (not heat-treated)  
**Instructions:** Specimen should be transported to the laboratory as soon as possible after collection. For longer storage, serum or plasma (separated from whole blood) may be frozen at ~20°C or below (no freeze-thaw) prior to transport.  
**Expected Value:** Genotype 1, Genotype 2, Genotype 3, Genotype 4, Genotype 5, Genotype 6   | No            |
| HERPES SIMPLEX VIRUS (HSV) CULTURE       | **Description:** Isolation of Herpes Simplex Virus (types 1 & 2) from clinical specimens  
**Specimen:** Oral or genital lesion swabs placed in viral transport media; biopsy or autopsy material placed in viral transport media; CSF collected in sterile container  
**Instructions:** All specimens must be transported using cold packs.  
If transport is delayed, specimens may be refrigerated at 2-8°C for up to 72 hours. Cotton or Dacron tipped swabs (DO NOT use swabs with calcium alginate or wood shafts)  
**Expected Value:** No evidence of HSV in culture   | Yes           |
| HERPES SIMPLEX VIRUS (HSV) ANTIBODY IgG  | **Description:** Detection of Herpes Simplex virus antibody by ELISA. The Immunoblot confirmatory test will automatically be run on specimens with positive screening results  
**Specimen:** 1-2 ml Serum  
**Instructions:** Refrigerate specimens at 2-8°C until transport  
**Expected Value:** Negative for Herpes Simplex Virus Antibody I and II   | Yes           |
| HIV-1 WESTERN BLOT                       | **Description:** Confirmation of HIV-1 screening results using Western Blot technology  
**Specimen:** 1-2 ml Serum  
**Instructions:** Refrigerate specimens at 2-8°C before transport. If storage time exceeds two weeks before transport, freeze serum or plasma (separated from whole blood) at -20°C or below.  
**Expected Value:** Nonreactive for HIV-1 by Western Blot   | Yes           |
| INFLUENZA CULTURE                        | **Description:** Isolation and subtyping of Influenza virus (type A or B) from clinical specimens  
**Specimen:** Throat or nasopharyngeal; Autopsy/biopsy specimens  
**Instructions:** Throat or nasopharyngeal: Flocked, cotton or Dacron tipped swabs (DO NOT use swabs with calcium alginate or wood shafts) placed in viral transport media; CSF collected in sterile container.  
Autopsy/biopsy specimens: Place in viral transport media  
Transport to laboratory within 24 hours using cold packs.  
**Expected Value:** No evidence of Influenza virus in culture   | Yes           |
| INFLUENZA RT-PCR                         | **Description:** Detection and characterization of Influenza virus (type A or B) from clinical specimens  
**Specimen:** Nasopharyngeal (NP) or nasal swabs placed in viral transport media and held at 2-8°C.  
**Instructions:** Collect the specimen using swabs with a synthetic tip (such as nylon or Dacron) and an aluminum or plastic shaft. Place the swab in viral transport media. Flocked NP swabs are the preferred specimen type. Transport to the laboratory within 72 hours using cold packs.  
**Expected Value:** Influenza virus RNA not detected   | Yes           |
<table>
<thead>
<tr>
<th>TEST</th>
<th>DESCRIPTION</th>
<th>TRANSPORT KIT?</th>
</tr>
</thead>
<tbody>
<tr>
<td>ISOSPORA</td>
<td><strong>DISCONTINUED 2016</strong></td>
<td>Yes</td>
</tr>
</tbody>
</table>
| LEGIONELLA CULTURE | **Description**: Selective media culture for Legionellae on buffered charcoal yeast extract (BCYE)  
|               | **Specimen**: Respiratory secretions ONLY (sputa, bronchial washing/brushing, pleural fluid, lung tissue, etc.) in sterile container.  
|               | **Instructions**: Refrigerate 2-8°C until transport. Specimens received >7 days from collection are unacceptable for testing.  
|               | **Expected Value**: No Legionella isolated                                    | No             |
| LEGIONELLA DFA | **Description**: Direct fluorescent antibody stain/smear for *Legionella pneumophila* types 1-6.  
|               | **Specimen**: Respiratory secretions ONLY (sputa, bronchial washing/brushing, pleural fluid, lung tissue, etc.) in sterile container.  
|               | **Instructions**: Refrigerate 2-8°C until transport. Specimens received >7 days from collection are unacceptable for testing.  
|               | **Expected Value**: No *Legionella pneumophila* types 1-6 detected            | No             |
| MEASLES (Rubeola) IgG | **Description**: Detection of Measles IgG antibody by ELISA  
|               | **Specimen**: 1-2 ml serum  
|               | **Instructions**: May be refrigerated at 2-8°C for up to 7 days. If storage time exceeds 7 days, freeze serum at -20°C or below  
|               | **Expected Value**: Negative for Measles IgG antibody                           | Yes            |
| MEASLES (Rubeola) IgM | **Description**: Detection of Measles IgM antibody by ELISA  
|               | **Specimen**: 1-2 ml serum  
|               | **Instructions**: Collect specimen ASAP after onset of symptoms. May be refrigerated at 2-8°C for up to 7 days. If storage time exceeds 7 days, freeze serum at -20°C or below  
|               | **Expected Value**: Negative for Measles IgM antibody                           | Yes            |
| MEASLES RT-PCR | **Description**: Detection of Measles virus by real time RT–PCR  
|               | **Specimen**: Throat swab in viral transport medium  
|               | **Instructions**: Refrigerate at 2-8°C until transport  
|               | **Expected Value**: Measles virus not detected                                 | Yes            |
| MERCURY      | **Description**: Quantitation of mercury (Hg) in blood by Inductively Coupled Plasma Mass Spectrometry (ICP-MS)  
|               | **Specimen**: 1 ml Whole Blood in EDTA Vacutainer tube  
|               | **Instructions**: Refrigerate specimens when not in transit. Do not freeze.  
|               | **Expected Value**: <15 ug/L                                                  | No             |
| MUMPS IgG    | **Description**: Detection of Mumps IgG antibody by ELISA  
|               | **Specimen**: 1-2 ml serum  
|               | **Instructions**: Specimens may be refrigerated at 2-8°C for up to 2 days. For longer storage, freeze at -20°C or below. Paired sera are required to confirm acute infection. Collect acute specimen as soon as possible after onset of symptoms but not later than seven days after onset. The convalescent specimen should be collected 14-21 days after the acute serum.  
<p>|               | <strong>Expected Value</strong>: Positive for Mumps IgG antibody                            | Yes            |</p>
<table>
<thead>
<tr>
<th>TEST</th>
<th>DESCRIPTION</th>
<th>TRANSPORT KIT?</th>
</tr>
</thead>
</table>
| MUMPS IgM        | **Description:** Detection of Mumps IgM antibody by ELISA  
|                  | **Specimen:** 1-2 ml serum                                                  | Yes            |
|                  | **Instructions:** Submit serum for testing as soon as possible after collection. Collect acute specimen between seven days and up to four weeks after onset of symptoms. Specimens may be refrigerated at 2-8°C. |
|                  | **Expected Value:** Negative for Mumps IgM antibody                         |                |
| MUMPS RT-PCR     | **Description:** Detection of Mumps virus by real time RT–PCR  
|                  | **Specimen:** Buccal swab in viral transport medium                          | Yes            |
|                  | **Instructions:** Refrigerate at 2-8°C until transport                       |                |
|                  | **Expected Value:** Mumps virus not detected                                |                |
| MYCOBACTERIOLOGY | **See TB - Acid-Fast Bacillus**                                              |                |
| MYCOLOGY         | **See Fungal Culture**                                                      |                |
| NOROVIRUS RT-PCR | **Description:** Detection of Norovirus by RT-PCR  
|                  | **Specimen:** Stool specimen in sterile container                           | No             |
|                  | **Instructions:** Refrigerate at 2-8°C until transport                       |                |
|                  | **Expected Value:** Norovirus not detected                                  |                |
| OVA & PARASITES  | **Description:** Direct and concentrated wet mounts, trichrome smear, Cryptosporidium and Giardia DFA. Note: Screen for Cyclospora, Isospora, Sarcocystis and Microsporidium NOT included  
|                  | **Specimen:** Fecal material in 10% formalin, Streck tissue fixative (STF), sodium acetate-acetic acid formalin (SAF) or Ecofix  
|                  | **Instructions:** Store and transport at ambient conditions                | Yes            |
|                  | **Expected Value:** No Ova or Parasites found                               |                |
| PERTUSSIS CULTURE| **Description:** Selective culture on Regan-Lowe (RL) media for Bordetella pertussis and Bordetella parapertussis  
|                  | **Specimen:** One nasopharyngeal calgiswab in Regan-Lowe transport for culture. Throat swabs are unacceptable.  
|                  | **Instructions:** Transport at ambient conditions. If transported within 24 hrs. store RL at room temp (20–25°C). If transported within 48–72 hrs. incubate at 34–37°C until shipment. If incubator is unavailable, refrigerate until shipment. Specimens received >7 days from collection are unacceptable for testing |
|                  | **Expected Value:** No Bordetellae found by culture                         | Yes            |
| PERTUSSIS PCR    | **Description:** Polymerase chain reaction for Bordetella pertussis, Bordetella parapertussis, and Bordetella holmesii  
|                  | **Specimen:** One nasopharyngeal polyester (DACRON) swabs for PCR in plain sterile plastic tube. Throat swabs are unacceptable.  
|                  | **Instructions:** Refrigerate PCR tubes with swabs until shipment. Transport at ambient conditions. Specimens received >7 days from collection are unacceptable for testing |
|                  | **Expected Value:** No Bordetellae found by PCR                            | Yes            |

**DISCONTINUED 2016**
<table>
<thead>
<tr>
<th>TEST</th>
<th>DESCRIPTION</th>
<th>TRANSPORT KIT?</th>
</tr>
</thead>
</table>
| RESPIRATORY PANEL          | Description: Multiplex amplified assay that detects 19 different respiratory pathogens (Adenovirus, Coronavirus 229E, Coronavirus HKU1, Coronavirus NL63, Coronavirus OC43, Human Metapneumovirus, Influenza A, Influenza A subtype H1, Influenza A subtype H3, Influenza A subtype H1-2009, Influenza B, Parainfluenza Virus 1, Parainfluenza Virus 2, Parainfluenza Virus 3, Parainfluenza Virus 4, Human Rhinovirus/Enterovirus, Respiratory Syncytial Virus, Bordetella pertussis, Chlamyphila pneumoniae, and Mycoplasma pneumoniae).  
Specimen: Nasopharyngeal Swab in Viral Transport Medium (VTM)  
Instructions: Place swabs in VTM immediately after collection. If storage is required, specimens in VTM can be held at room temperature (18–30°C) for up to 4 hours, or at refrigerator temperature (2-8°C) for up to 3 days. Transport to laboratory as soon as possible after collection in order to ensure testing within 3 days of collection  
Expected Value: None Detected | Yes |
| RPR (ROUTINE), QUALITATIVE  | Description: The Rapid Plasma Reagin (RPR) Card Test is a nontreponemal, macrofloculation test for syphilis used qualitatively for screening. All reactive RPR's are titred to an endpoint (see RPR, quantitative)  
Note: Patients should not have eaten for one hour prior to collection.  
Specimen: 1 ml serum  
Instructions: Refrigerate specimens at 2-8°C until transport  
Note: Avoid hemolysis, which may interfere with the test.  
Expected Value: Nonreactive | Yes |
| RPR, QUANTITATIVE           | Description: The Rapid Plasma Reagin (RPR) Card Test is a nontreponemal, macrofloculation test for syphilis quantitatively to monitor treatment response to the disease. All reactive RPR's are titred to an endpoint  
An endpoint titer is determined by performing the RPR test (see above) on a series of serum dilutions  
Specimen: 1 ml serum  
Instructions: Refrigerate specimens at 2-8°C until transport  
Expected Value: <1:16 or Endpoint dilution | Yes |
| RUBELLA IgG                 | Description: Detection of Rubella IgG antibody by ELISA  
Specimen: 1-2 ml serum  
Instructions: Specimen may be refrigerated at 2-8°C for up to 5 days until transport. To confirm acute infection, acute serum should be collected within 7 days of onset; convalescent serum 2-3 weeks after acute serum. Acute serum may be frozen at -20°C or below until convalescent serum is collected  
Expected Value: Positive or Negative for Rubella IgG antibody | Yes |
| RUBELLA IgM                 | Description: Detection of Rubella IgM antibody by ELISA  
Specimen: 1-2 ml serum  
Instructions: Collect specimen 8-21 days post onset of rash. If suspected congenital infection, a neonatal serum specimen should be collected as soon as possible following birth. Specimen may be refrigerated at 2-8°C for up to 5 days prior to transport  
Expected Value: Negative for Rubella IgM antibody | Yes |
<p>| RUBEOLA                     | See Measles                                                                                                                                                                                               | Yes |</p>
<table>
<thead>
<tr>
<th>TEST</th>
<th>DESCRIPTION</th>
<th>TRANSPORT KIT?</th>
</tr>
</thead>
<tbody>
<tr>
<td>ST. LOUIS ENCEPHALITIS (SLE) SEROLOGY</td>
<td>See Arbovirus Serology Panel</td>
<td>Yes</td>
</tr>
<tr>
<td>SARCOCYSTIS</td>
<td>DISCONTINUED 2016</td>
<td></td>
</tr>
<tr>
<td>SEROGROUPING</td>
<td><strong>Description:</strong> Latex or antiserum “somatic grouping” of isolates. Species available: <em>Neisseria meningitidis</em>, <em>Haemophilus influenzae</em>, Beta hemolytic Streptococci, <em>Shigella</em>, <em>Vibrio cholerae</em>&lt;br&gt;&lt;br&gt;<strong>Specimen:</strong> Pure isolate in supportive environment/media&lt;br&gt;&lt;br&gt;<strong>Instructions:</strong> Transport at ambient conditions. Tubed media must be used for transport by common carrier&lt;br&gt;&lt;br&gt;<strong>Expected Value:</strong> Genus, species and serogroup of organism</td>
<td>No</td>
</tr>
<tr>
<td>SHIGA-LIKE TOXINS (SLT)</td>
<td><strong>Description:</strong> EIA test to detect SLT-I&amp;II; see Hemorrhagic <em>E.coli Screen/Confirmation.</em>&lt;br&gt;Stool in fecal transport (Cary-Blair or equivalent), stool in MAC or GN broth or an <em>Escherichia coli</em> isolate.&lt;br&gt;&lt;br&gt;<strong>Instructions:</strong> Transport stool in fecal transport at ambient conditions; must receive within 3 days of collection. Stool in broth must be received within 7 days. Refrigerate if delay in transport &gt;24 hours.&lt;br&gt;&lt;br&gt;<strong>Expected Value:</strong> Negative for Shiga-like toxins</td>
<td>Yes</td>
</tr>
<tr>
<td>SMEAR (DIRECT)</td>
<td><strong>Description:</strong> Stained smear appropriate for organism sought (e.g., gram stain, Calcofluor White/KOH, Acid Fast, Modified Acid Fast, Loeffler Methylene Blue, etc.)&lt;br&gt;&lt;br&gt;<strong>Specimen:</strong> Prepared smears (preferably 2 smears) for staining&lt;br&gt;&lt;br&gt;<strong>Instructions:</strong> Transport at ambient conditions in slide holder. Specify organism/stain desired&lt;br&gt;&lt;br&gt;<strong>Expected Value:</strong> Absence or presence of organism requested</td>
<td>No</td>
</tr>
<tr>
<td>SYPHILIS</td>
<td>See TP-PA; RPR – Qualitative, Quantitative; VDRL</td>
<td>Yes</td>
</tr>
<tr>
<td>TB - Acid-Fast Bacillus (AFB) CULTURE, and IDENTIFICATION</td>
<td><strong>Description:</strong> Detection and identification of <em>Mycobacterium</em> species&lt;br&gt;&lt;br&gt;<strong>Specimen:</strong> Respiratory:&lt;br&gt;– Sputum – 5 ml coughed or induced&lt;br&gt;– Bronchial wash – 5 ml&lt;br&gt;&lt;br&gt;Tissue: Skin scrapings, biopsy, necropsy.&lt;br&gt;Swabs are discouraged&lt;br&gt;&lt;br&gt;Body fluids:&lt;br&gt;– Blood - 5 ml SPS tube only (yellow cap)&lt;br&gt;– CSF - 2 ml minimum&lt;br&gt;– Urine - 10 - 50 ml first AM void&lt;br&gt;– Gastric lavage - 5 - 10 ml immediately neutralized with 100 mg of sodium carbonate&lt;br&gt;– Other fluids - 5 ml minimum&lt;br&gt;&lt;br&gt;<strong>Instructions:</strong> Refrigerate until transport. Transport at ambient conditions&lt;br&gt;&lt;br&gt;<strong>Expected Value:</strong> No <em>Mycobacterium</em> species detected or isolated</td>
<td>Yes</td>
</tr>
<tr>
<td>TEST</td>
<td>DESCRIPTION</td>
<td>TRANSPORT KIT?</td>
</tr>
<tr>
<td>------</td>
<td>-------------</td>
<td>---------------</td>
</tr>
</tbody>
</table>
| **TB – Acid-Fast Bacillus (AFB) SMEAR only*** | **Description:** Stain for Acid-Fast Bacilli - for screening purposes only  
**Specimen:** Respiratory: Sputum – 5 ml coughed or induced  
Bronchial wash – 5 ml  
Tissue: Skin scrapings, biopsy, necropsy; Swabs are discouraged  
**Body fluids:** Blood - 5 ml SPS tube only (yellow cap)  
CSF - 2 ml minimum  
Urine - 10 - 50 ml first AM void  
Gastric lavage - 5 - 10 ml immediately neutralized with 100 mg of sodium carbonate  
Other fluids - 5 ml minimum  
**InSTRUCTIONS:** Refrigerate until transport. Transport at ambient conditions  
**Expected Value:** No AFB seen  
*PHL does not recommend the diagnosis of Mycobacteria on “smear only” results. Cultures are recommended for recovery of the microorganism. | Yes |
| **TB – ISOLATE MYCOBACTERIA IDENTIFICATION** | **Description:** Identification of Mycobacteria using biochemicals and/or DNA probes  
**Specimen:** Mycobacterium isolate on Lowenstein-Jensen (L-J)  
**Instructions:** Transport at ambient conditions  
**Expected Value:** Identification of organism | No |
| **TB - MYCOBACTERIUM TUBERCULOSIS NAA DIRECT TEST** | **Description:** Detection of *M. tuberculosis* complex rRNA  
**Specimen:** Non-bloody respiratory specimen in sterile container  
**Instructions:** Refrigerate at 2–8°C until transport  
**Expected Value:** *Mycobacterium tuberculosis* complex rRNA not detected | No |
| **TB - MYCOBACTERIUM TUBERCULOSIS SUSCEPTIBILITIES** | **Description:** *Mycobacterium tuberculosis* susceptibility testing  
**Specimen:** *Mycobacterium tuberculosis* isolate on L-J media  
**Instructions:** Transport at ambient conditions  
**Expected Value:** Sensitive to Streptomycin, Isoniazid (INH), Rifampin, Ethambutol, Pyrazinamide (PZA) (determined by % resistance) | |
| **TP-PA (SYPHILIS)** | **Description:** *Treponema pallidum* passive particle agglutination to detect the presence of antibody to *T. pallidum* in serum. **This test is reserved primarily for confirming reactive nontreponemal tests**  
**Specimen:** 1 ml serum or plasma  
**Instructions:** Specimens may be refrigerated at 2-8°C for up to 5 days prior to transport. For longer storage, freeze at -20°C or below. Plasma specimens must be tested within 48 hours of collection. **Note:** Hemolysis may interfere with the test.  
**Expected value:** Nonreactive | Yes |
| **VARICELLA IgG** | **Description:** Detection of Varicella IgG antibody by ELISA  
**Specimen:** 1-2 ml serum  
**Instructions:** Specimen may be refrigerated at 2-8°C for up to 7 days prior to transport. For longer storage, freeze at -20°C or below. To confirm acute infection, acute serum should be collected as soon as possible after onset of symptoms; convalescent serum 14-21 days after collection of acute serum  
**Expected Value:** Positive for Varicella IgG antibody | Yes |
<table>
<thead>
<tr>
<th>TEST</th>
<th>DESCRIPTION</th>
<th>TRANSPORT KIT?</th>
</tr>
</thead>
</table>
| VDRL                     | **Description:** Venereal Disease Research Laboratory slide test-qualitative and quantitative. The VDRL slide test is a nontreponemal, microflocculation test for syphilis used for screening and observing response to treatment. All reactive VDRL's are titered to an endpoint. **The PHL performs the VDRL slide test on CSF only.**  
**Specimen:** 1 ml aseptically collected CSF  
**Instructions:** Refrigerate specimen at 2-8°C until transport. Do not freeze or add preservative  
**Expected Value:** Nonreactive                                                                 | Yes            |
| VIBRIO SCREEN            | **Description:** Fecal specimen screened for Vibrio species.  
**Specimen:** Fecal specimen in fecal transport (Cary Blair or equivalent).  
**Instructions:** Transport at ambient conditions. Must be received within 3 days of collection. Refrigerate if delay in transport >24 hours.  
**Expected Value:** No Vibrio isolated                                                                 | Yes            |
| VIRAL CULTURE -          | **Description:** Isolation and identification of viruses from clinical specimens. Please specify virus suspected  
**Specimen:** Nasopharyngeal, throat, cervical, urethral, rectal, wound/lesion/ulcer swab - in viral transport medium;  
CSF: Minimum 0.5 ml in sterile tube  
Stool: pea-sized sample in viral transport medium  
Urine: 1-3 ml  
**Instructions:** Refrigerate at 2-8°C until transport. Specimens should be transported to laboratory within 24 hours of collection using cold packs. Cotton or Dacron tipped swabs **DO NOT** use swabs with calcium alginate or wood shafts  
**Expected Value:** No viruses isolated in culture                                                                 | Yes            |
|  Enterovirus             |                                                                 |                |
|  Herpes                  |                                                                 |                |
|  Mumps                   |                                                                 |                |
|  Respiratory             |                                                                 |                |
|  Varicella-Zoster        |                                                                 |                |
|  Other                   |                                                                 |                |
| VIRAL RESPIRATORY PANEL -| **Description:** Identification of Influenza A; Influenza B; Parainfluenza 1, 2, & 3; Respiratory Syncytial Virus (RSV); and Adenovirus  
**Specimen:** Nasopharyngeal or throat swab - place in 1-2 ml viral transport media  
Throat wash - in 3-5 ml sterile saline  
Nasal wash - in 2-3 ml sterile saline  
Nasopharyngeal wash - 3-7 ml  
Nasopharyngeal aspirate - 0.2-0.8 ml  
Specimens must be transported to laboratory within 24 hours of collection using cold packs. Cotton or Dacron tipped swabs **DO NOT** use swabs with calcium alginate or wood shafts  
**Instructions:**  
**Expected Value:** No evidence of Influenza A, B; Parainfluenza 1, 2, 3; RSV; or Adenovirus in culture                                                                 | Yes            |
|  Culture                 |                                                                 |                |
| WATER ANALYSIS           | For Water Analysis Laboratory services, refer to: [http://des.nh.gov/organization/commissioner/lsu/index.htm](http://des.nh.gov/organization/commissioner/lsu/index.htm) or call 603-271-3445                                                                 | Yes            |
| WEST NILE VIRUS (WNV)    | See Arbovirus Serology Panel                                                                 | Yes            |
| YEAST ISOLATE IDENTIFICATION | **Description:** Identification of fungi to genus; species level if appropriate  
Isolate (preferably pure) on supportive media  
**Specimen:** Samples must be in tubed media for common carrier transport. Transport and hold at ambient conditions.  
**Expected Value:** Genus and species of organism identification                                                                 | Yes            |
| YERSINIA SCREEN          | **Description:** Fecal specimen screened for Yersinia species.  
**Specimen:** Fecal specimen in fecal transport (Cary Blair or equivalent).  
**Instructions:** Transport at ambient conditions. Must receive within 3 days of collection. Refrigerate if delay in transport >24 hours.  
**Expected Value:** No Yersinia isolated                                                                 | Yes            |
Notes:

- To order Transport Kits call 603-271-4605
- For more information call the NH Public Health Laboratories at 603-271-4661
- Fax number: 603-271-4783
- For collection instructions and test requisitions for clinical testing, please refer to our website at:
  
  http://www.dhhs.nh.gov/dphs/lab/labrequisitions.htm

TESTING CHANGES

- As of September 2012, Cytomegalovirus Culture was discontinued
- 2015 – HIV changes – New Ag/Ab Combo for routine testing, HIV1/2/Group O for decedent or cadaver testing only; discontinued HIV Western Blot
  - New tests: Chikungunya, Respiratory panel
  - Chemistry change: Mercury urine discontinued, blood mercury added
- 2016 – Discontinued tests: Ova & Parasites, Isospora, Giardia/Cryptosporidium, Sarcocystis